

Macs-based enrichment of untouched CD4+CD8+ thymocytes for biochemistry

CD53 depletion protocol

- 1) Dissect thymuses into Medium-199 (+ 2% FCS, 1x Pen/Strep, 1X Hepes)
- 2) Make single cell suspensions
- 3) Stain thymocytes in 0.5 – 1 ml MACS running buffer (DPBS, 0.5% BSA, 2mM EDTA)
with anti-CD53 (Pharmingen cat # 559364 for 15 min on ice
50ul per 2E8 cells, 75ul for TCR transgenics)
- 4) Wash with 10-20X volume of MACS running buffer.
- 5) Aspirate buffer completely (important!)
- 6) Stain thymocytes in 0.5 – 1 ml MACS running buffer 15 min on ice
with anti-rat IgG-biotin (Jackson Immunoresearch cat # 112-065-143
100ul per 2E8 cells, 150ul for TCR transgenics)

**OR: with anti-rat IgM-biotin
20 ul per 2E8 cells**
- 7) Wash with 10-20X volume of MACS running buffer.
- 8) Add Miltenyi Biotech anti-biotin beads and stain on ice 15 min
(100ul per 2E8 cells), with intermittent mixing.
- 9) Wash with 10-20X volume of MACS running buffer.
- 10) Resuspend in 2 ml MACS running buffer and separate on MACS column.
- 11) Assess purity by flow cytometry.

